

# Defining validation criteria for a primary jejunum and primary hepatocyte dual-organ MPS: a promising tool for more predictive studies of human drug ADME and oral bioavailability

Yassen Abbas<sup>1</sup>, Hailey Sze<sup>1</sup>, Christiana Skarlatopoulou<sup>1</sup>, Ashley A. Spreen<sup>2</sup>, Elizabeth M. Boazak<sup>2</sup>, William R. Thelin<sup>2</sup> and Tomasz Kostrzewski<sup>1</sup>

1. CN Bio Innovations, 332 Cambridge Science Park, Cambridge, UK. 2. Altis Biosystems, 6 Davis Drive, Durham, NC27709, USA

## 1. Abstract

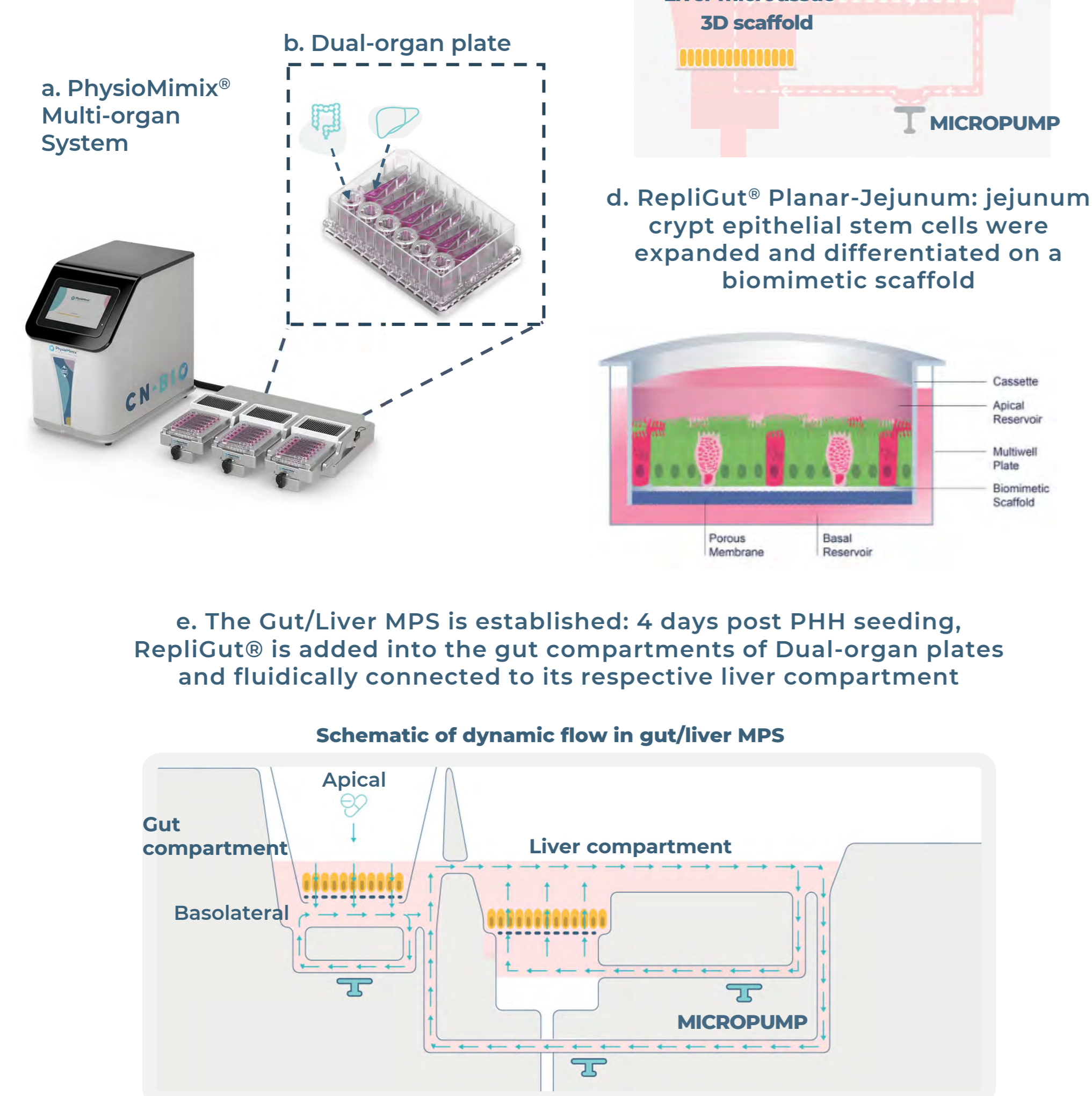
Efforts to improve the *in vitro* to *in vivo* translation of drug efficacy and safety data has led to the emergence of more physiologically-relevant microphysiological systems (MPS) that consist of multiple fluidically linked organs<sup>1</sup>. Here, we combine two established and well-characterized human MPSs, the RepliGut<sup>®</sup> Jejunum and PhysioMimix<sup>®</sup> Liver MPS, in an interconnected dual-organ MPS, to create a Gut/Liver system capable of profiling oral bioavailability.

Defining a cell validation criteria is important to ensure donor lots of liver and intestinal cells are functional in co-culture and meet the threshold for regulatory requirements and market adoption<sup>2</sup>.

Here, we define cell validation criteria whereby liver and intestinal cells are first validated separately, then as a functional, fluidically coupled co-culture system. We use a test compound to ensure the cells in co-culture are metabolically suitable for ADME studies. Finally, through real-world drug examples we demonstrate how the Gut/Liver MPS can be used to provide a mechanistic understanding of a drug's oral bioavailability *in vitro*.

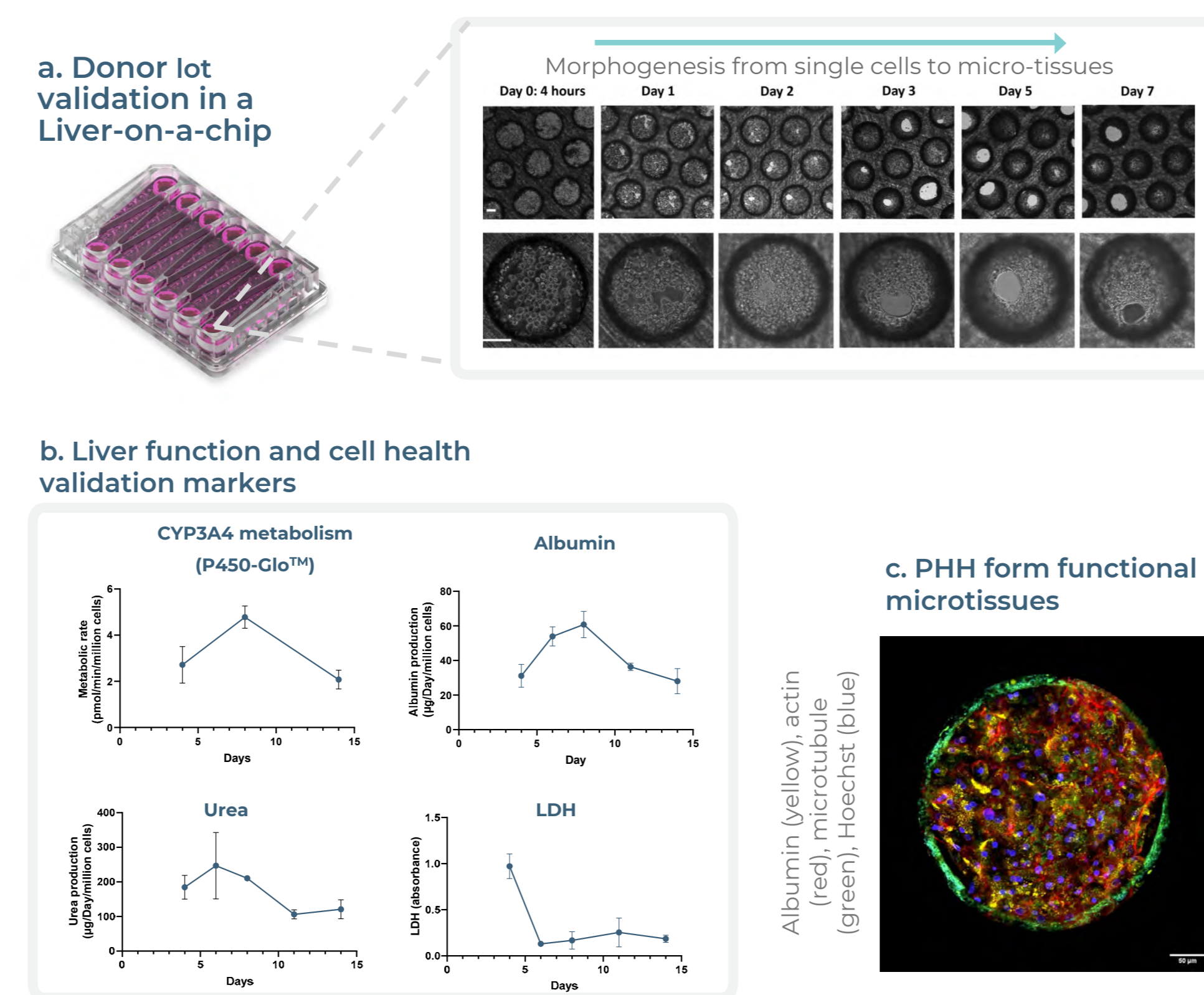
## 2. Methods

Fig 1. Establishment of the Gut/Liver MPS



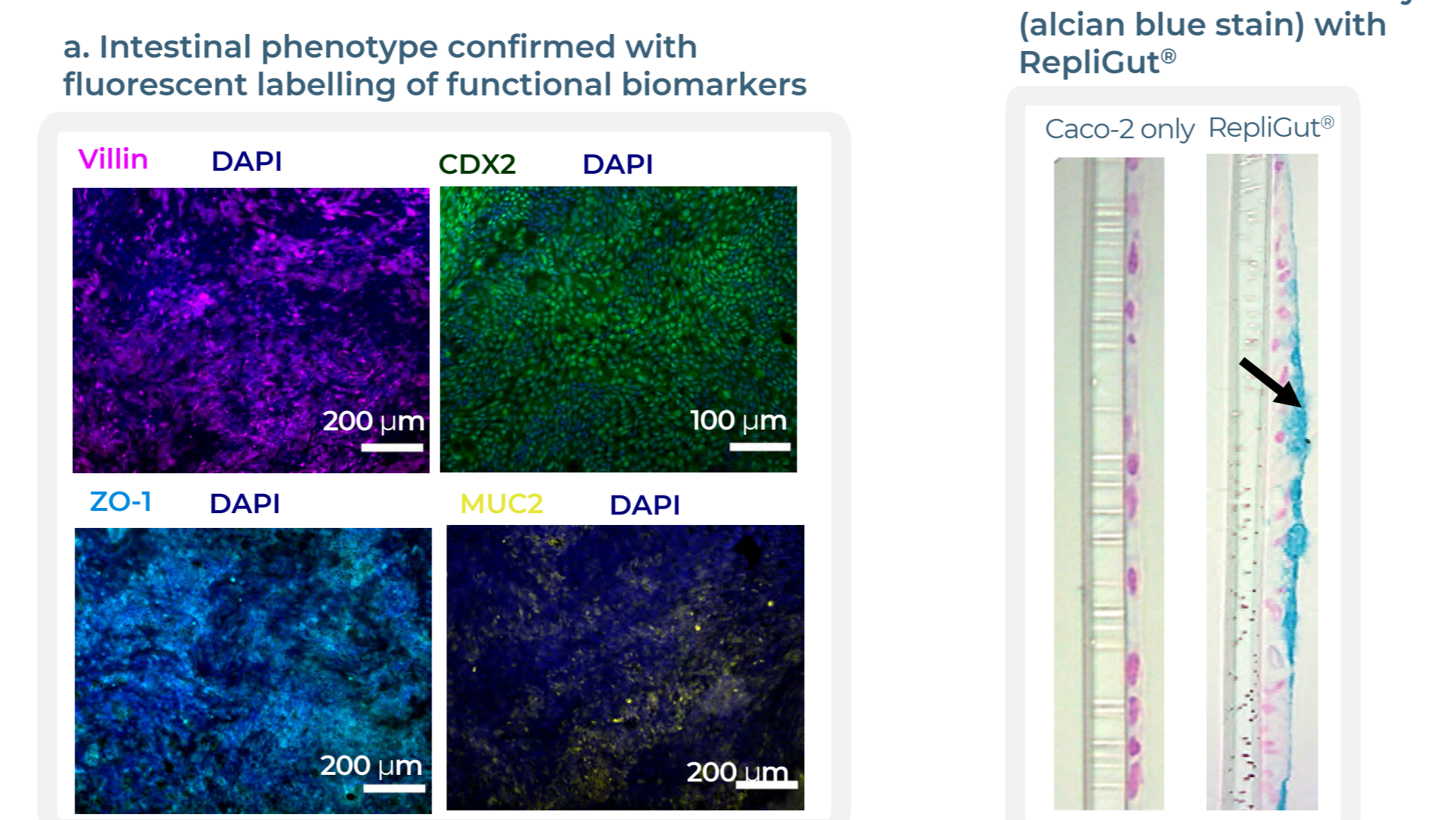
## 3. Results

Fig 2. Validation of PHH donor first in a Liver-on-a-chip to confirm metabolically functional tissue.

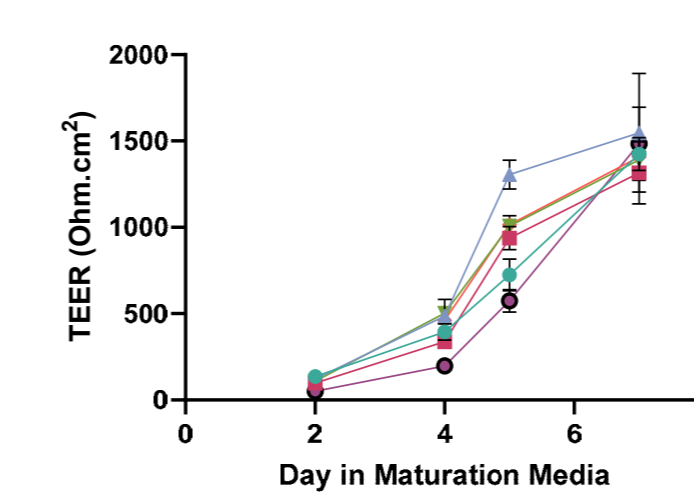


- PHH donors are pre-selected for a Gut/Liver MPS by first validating in a Liver-on-a-chip over a 14-day experiment.
- Functionality is assessed by CYP3A4 activity, albumin and urea production. Lactate dehydrogenase (LDH), a cell health marker, peaks at day 4 after the formation of microtissues, then remains low over the course of the experiment.

Fig 3. Validation of RepliGut<sup>®</sup> donor to confirm tissue functionality and reproducibility.



c. Reproducible TEER profiles from a single donor and cell lot in static



d. Two cell lots from the same donor show comparable TEER profiles in static

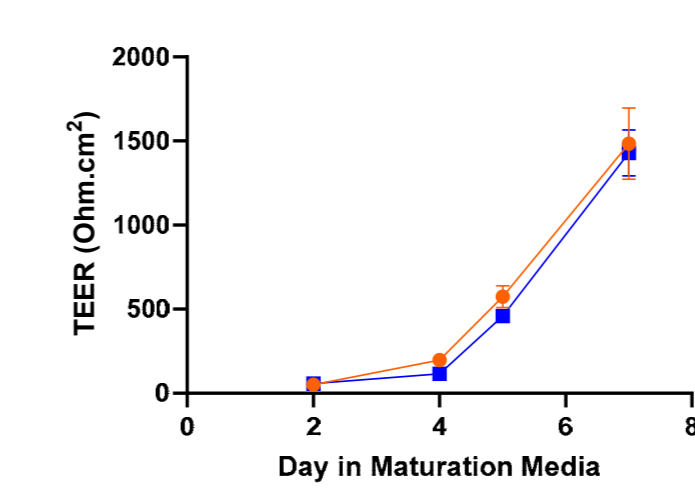


Fig 4. PHH functionality is maintained for at least 48 h in co-culture with RepliGut<sup>®</sup>

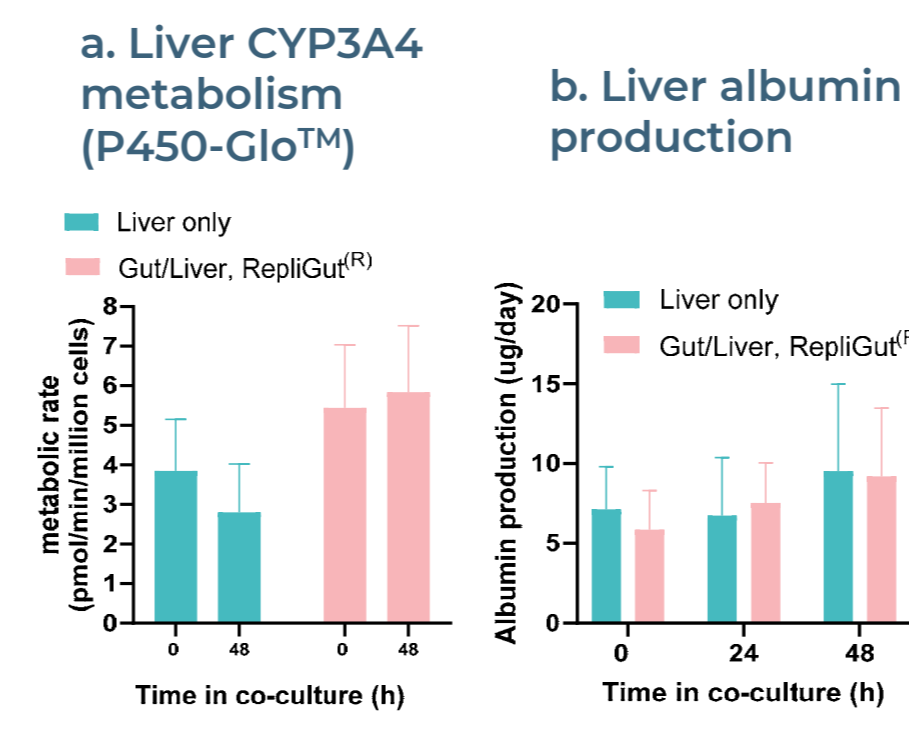


Fig 5. Intestinal barrier is maintained in co-culture with liver microtissues

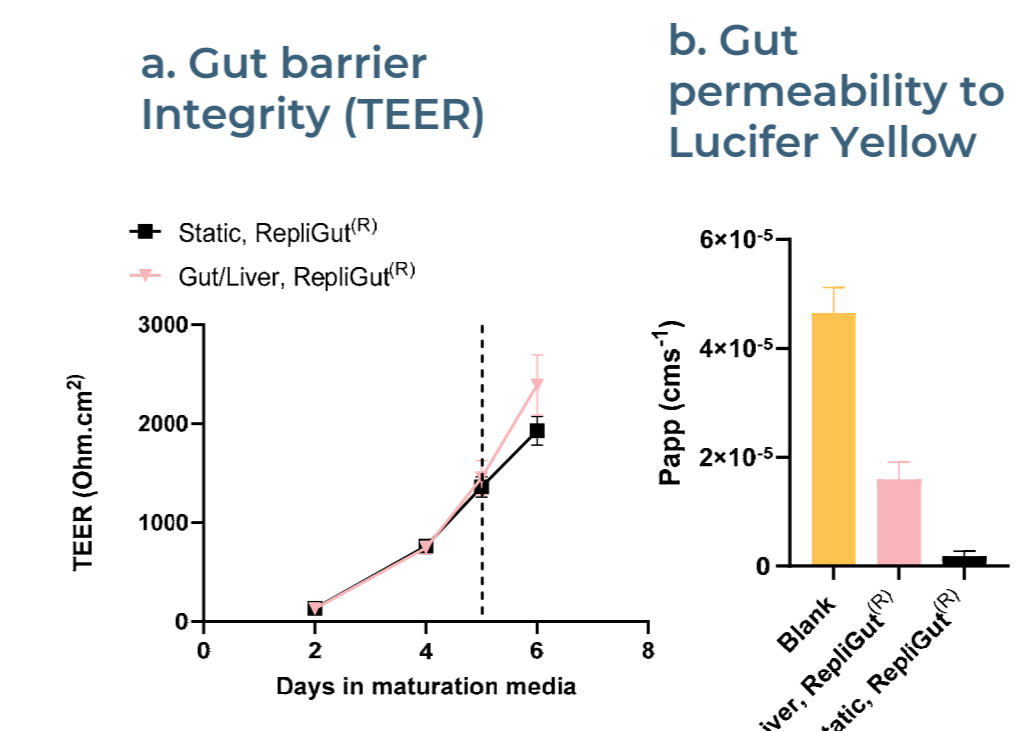


Fig 6. Validation with ADME test compound, 7-hydroxycoumarin (7-HC) to confirm intestinal absorption and hepatic metabolism in gut only, liver only and Gut/Liver MPS models.

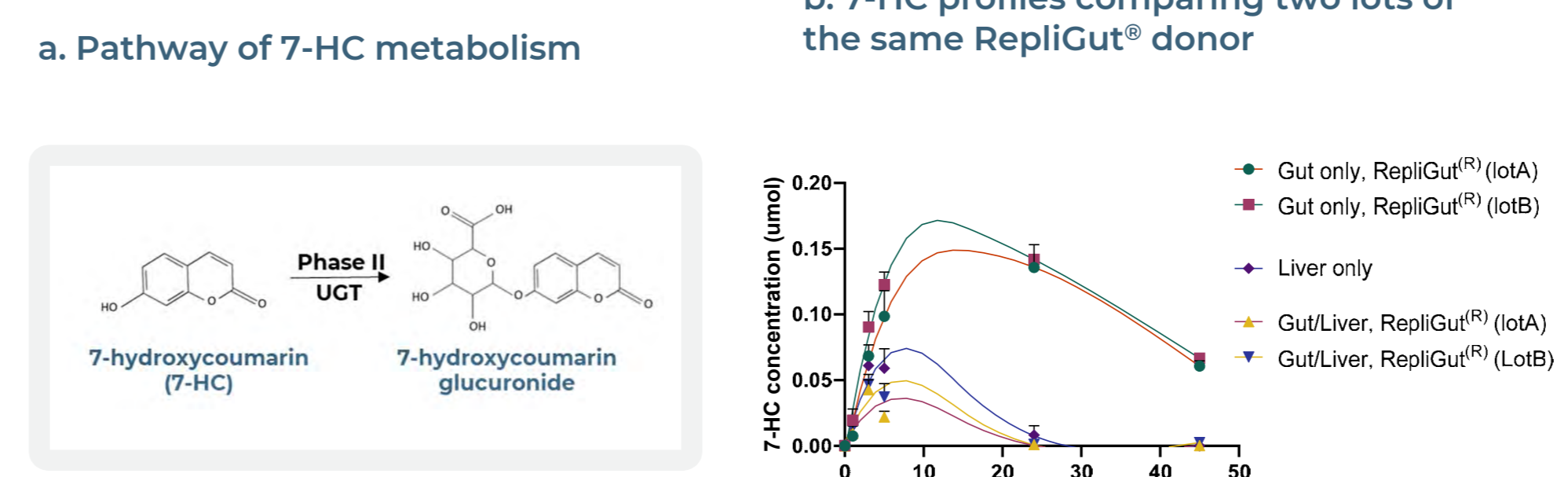


Fig 7. Case studies: profiling bioavailability on carboxylesterase (CES) mediated compounds.

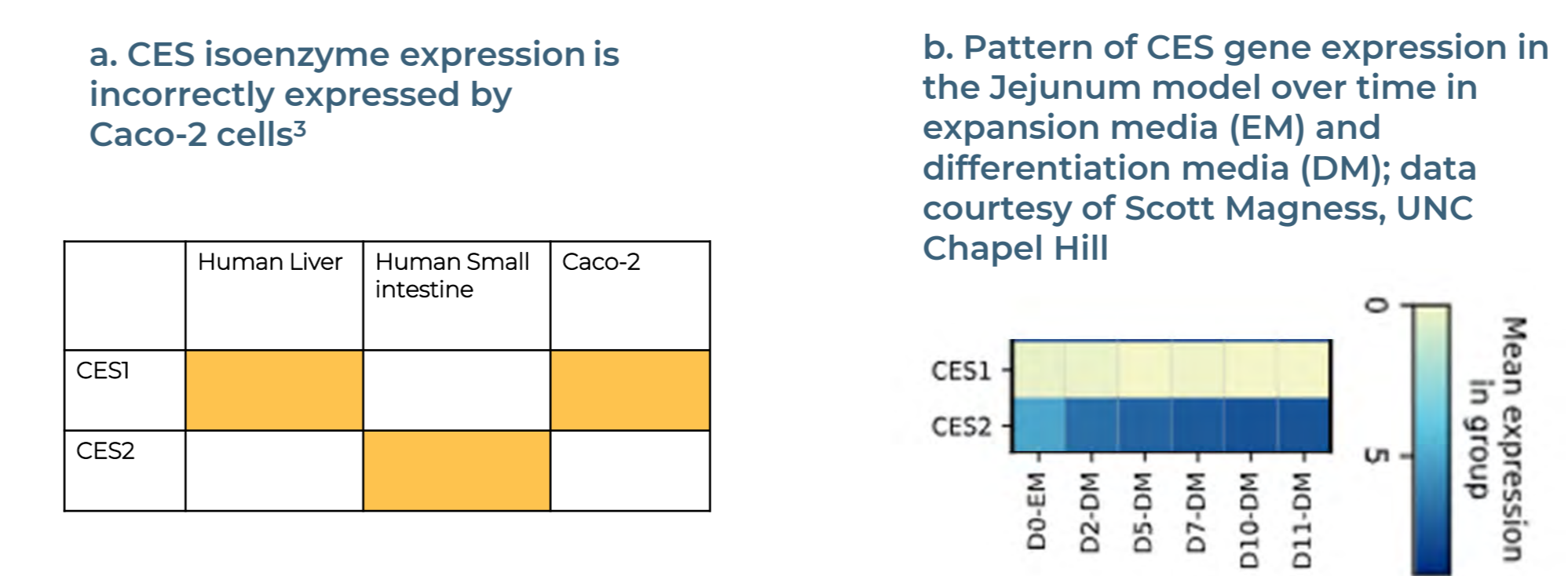


Fig 8. Case study 1, Enalapril: greater resistance to intestinal clearance observed in primary cell Gut/Liver MPS, correlates with isoenzyme expression in the human intestine

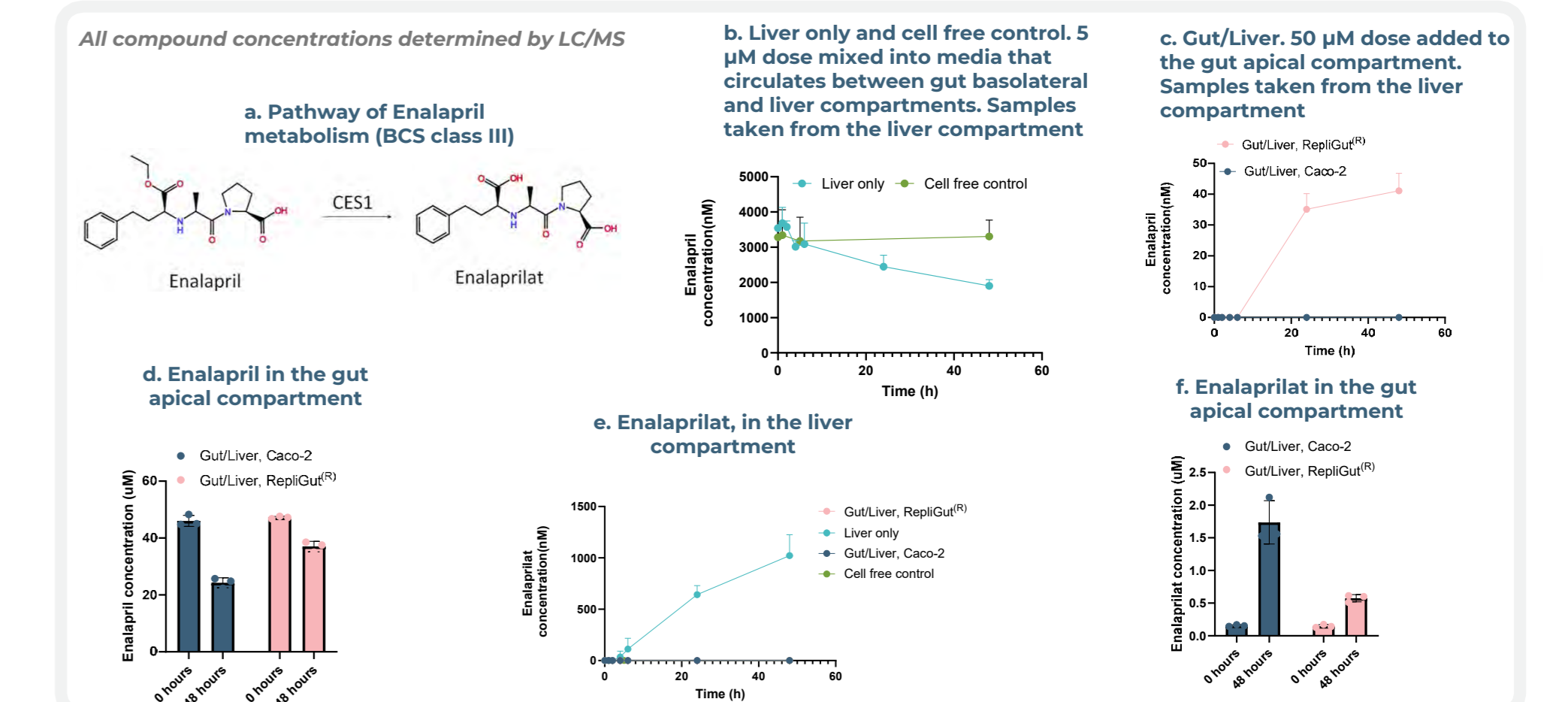
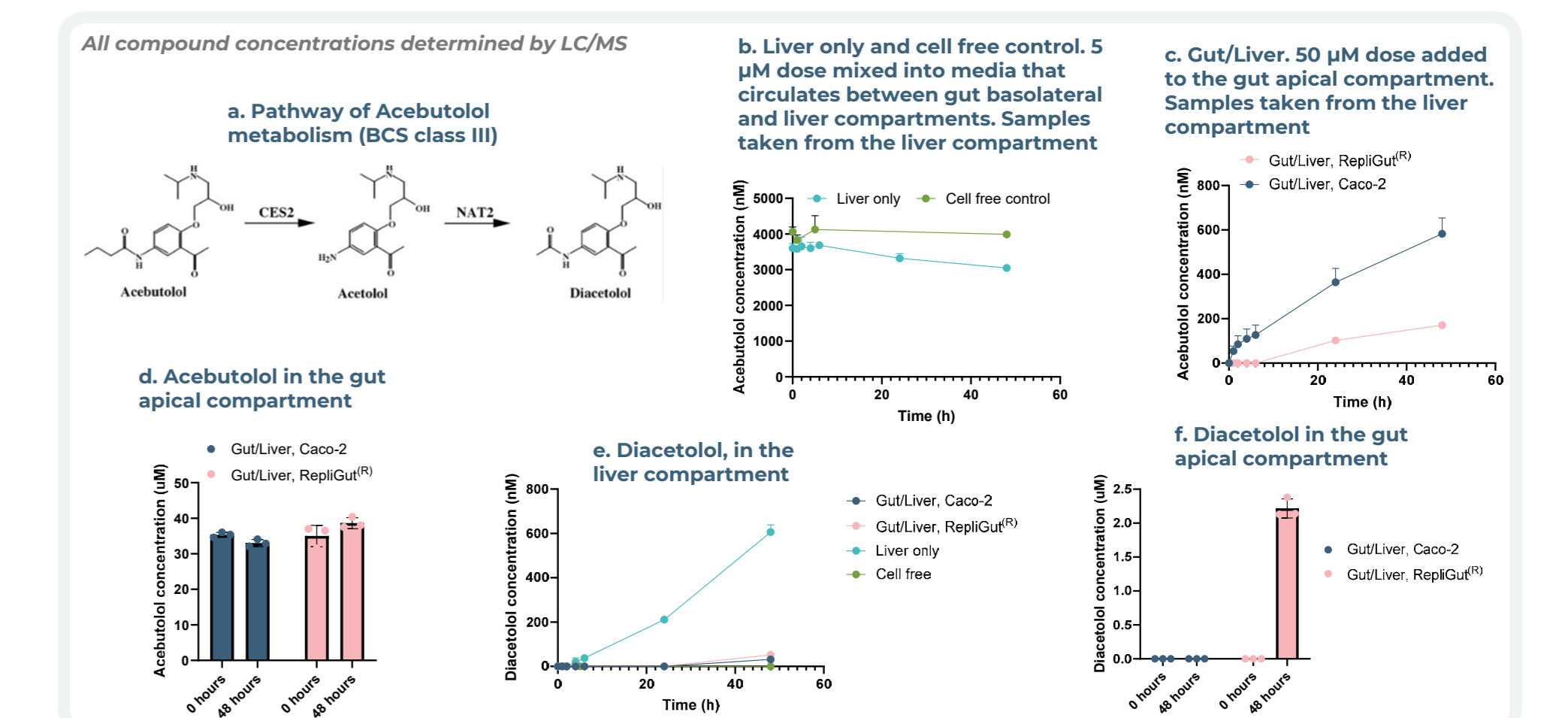


Fig 9. Case study 2, Acebutolol: the combination of intestinal metabolism and low permeability contribute to its low oral bioavailability. This is observed in the primary Gut/Liver MPS and correlates with acebutolol's bioavailability in humans



## 4. Conclusion

- The Gut/Liver MPS is a pre-clinical assay designed to profile human oral bioavailability *in vitro*.
- We provide validation criteria to ensure donor lots of liver and intestinal cells are functional in co-culture and are metabolically suitable for ADME studies.
- Through real-world drug examples, we demonstrate the assays utility to provide drug developers with a mechanistic understanding of bioavailability *in vitro*, allowing the progression of the most promising drug candidates.

1. C. D. Edgington et al., Sci. Reports 2018 8: 1-18 (2018).
2. T.K. Baker et al., Drug Metab. Dispos 52, 198-209 (2024).
3. T. Imai, et al, Drug Metab. Dispos. 33, 1185-1190 (2005).

